

Antibiotics

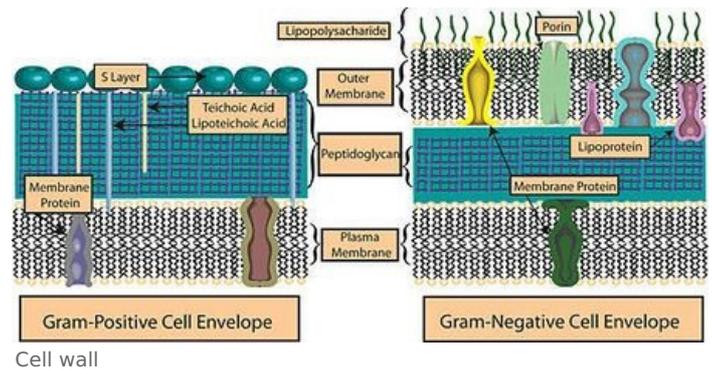
Antibiotics are substances that **inhibit the growth (multiplication) of microorganisms (bacteriostatic) or kill them (bactericidal)**. They are produced by bacteria or fungi. Their semisynthetic derivatives are also effective. More broadly, they include other antimicrobial agents, i.e. chemotherapeutics (synthetic substances) - sulphonamides and quinolones.



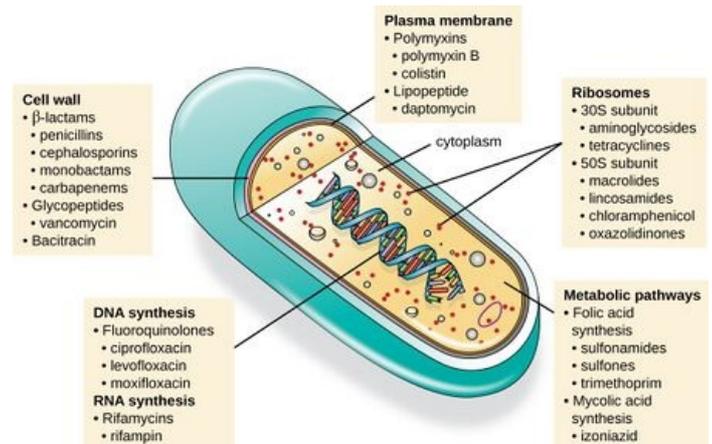
Antibiotics

The division between **bacteriostatic** and **bactericidal** is not entirely accurate, as many bacteriostatic antibiotics are also bactericidal in higher concentrations (chloramphenicol in meningococcal infection). Conversely, some bactericidal antibiotics do not kill certain bacteria even at high concentrations (penicillin G for *Enterococci*). Another aspect is the classification of antibiotics into groups according to their pharmacotherapeutic effect: antistaphylococcal, antipseudomonad, antianaerobic, antituberculous, etc.

Bakteriostatic	Baktericidal
<ul style="list-style-type: none"> ▪ Makrolide ▪ Tetracyclines ▪ Chloramphenicol ▪ Sulfonamides ▪ Trimethoprim ▪ Linkosamides ▪ Etambutol ▪ Nitrofurantoin 	<ul style="list-style-type: none"> ▪ Beta-lactam antibiotics <ul style="list-style-type: none"> ▪ Penicillins ▪ Cephalosporins ▪ Monobaktams ▪ carbapenems ▪ Aminoglykosides ▪ Peptides <ul style="list-style-type: none"> ▪ bacitracin ▪ colistin ▪ polymyxins ▪ Isoniazid ▪ Metronidazole ▪ nitrofurantoin ▪ Pyrazinamide ▪ Quinolones ▪ Rifampicin ▪ Glykopeptides <ul style="list-style-type: none"> ▪ Vankomycin ▪ Teikoplanin



Cell wall



Mechanism of action of antibiotics

Mechanisms of action

The basic requirement for antibiotic therapy is optimal antimicrobial effect with minimal toxicity to the host, i.e. high selectivity of action. This can be achieved in several ways:

1. by affecting structures or enzymatic processes specific only to microorganisms (cell wall synthesis),
2. accumulation of the substance in the microorganism using specific transport mechanisms (possible with tetracyclines).

Classification of antibiotics by mechanism of action

Inhibition of cell wall synthesis	Penicillins, cephalosporins, monobactams, carbapenems, vancomycin, bacitracin
Dysfunction of the cytoplasmic membrane	Amphotericin B, azoles, polyenes, polymyxins
Inhibition of protein synthesis	Aminoglycosides, chloramphenicol, macrolides, tetracyclines, lincomycin
Inhibition of nucleic acid synthesis	Sulfonamides, trimethoprim, quinolones, rifampicin, pyrimethamine

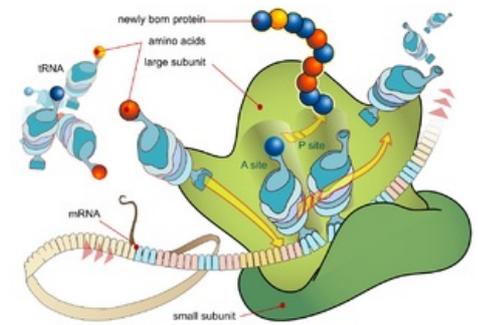
Inhibition of cell wall synthesis

Bacteria, unlike animal cells, have a cytoplasmic membrane covered by a cell wall that varies in structure, thickness and quality from strain to strain. The structure of the cell wall determines the Gram staining pattern and distinguishes **Gram-positive** and **Gram-negative** microorganisms.

The cell wall is essential for the survival of the microbe. It maintains its shape and ensures an optimal intracellular environment (high intracellular pressure). Damage to it (e.g. by lysol, detergents) or inhibition of the formation of any of its components leads to a breakdown of its function, even to cell lysis. This is particularly possible in grampositives.

Gram-positive microbes have a cell wall composed of a **thick layer of interconnected peptidoglycan chains**. The base of the peptidoglycan is a polysaccharide chain consisting of **N-acetylglucosamine** and **N-acetylmuramic acid**, to which is attached a polypeptide chain terminated by the 2 amino acids D-alanyl-D-alanine (D-Ala-D-Ala). The compact barrier is formed by cleavage of the terminal D-alanine and linking the rest of the polypeptide chain to the chain of the adjacent peptidoglycan - transpeptidation. This reaction is catalyzed by an integral penicillin binding protein (PBP). The synthesis of free peptidoglycan takes place on the inner side of the cytoplasmic membrane, transpeptidation on the outer side.

The cell wall of **Gram-negative** microorganisms is different. It consists of a **layer of peptidoglycans** (weaker than in gram-positives) supplemented by an **outer phospholipid membrane** that may prevent the penetration of hydrophilic antibiotics (penicillin G) to the binding protein. Hydrophilic antibiotics affect Gram-negative bacteria only if they are able to penetrate the transmembrane pores (porins) of the outer membrane (e.g. ampicillin, amoxicillin).



Synthesis of protein on ribosomes (translation)

Dysfunction of the cytoplasmic membrane

The ability to violate the integrity of the cytoplasmic membrane of bacteria (Gram-negatives due to polymyxins) or fungi (due to polyenes, azoles, amphotericin B) leads to the leakage of endogenous substances extracellularly and the subsequent death of the microorganism.

Inhibition of protein synthesis

The selectivity of the effect is due to inhibition of protein synthesis by bacterial 70S ribosomes.

Normal protein synthesis on ribosomes (translation) occurs in several steps. The stimulus for initiation is the binding of the mRNA initiation codon to the smaller subunit of the **ribosome (30S)**. The tRNA with a complementary triplet of bases (anticodon) is extracted from the free-living activated amino acids (bound to the tRNA) and binds to the mRNA initiation codon, forming the so-called initiation complex. In the next step, the acceptor site A (aminoacyl) on the larger subunit of the **ribosome (50S)** is released. This is bound by a tRNA that carries an anticodon complementary to another triplet on the mRNA. The enzyme peptidyltransferase is part of the larger subunit. It catalyzes the cleavage of the initial (N-terminal) amino acid from its tRNA and transfer to the next activated amino acid bound at the A site to form a peptide bond. The initial tRNA is released from the ribosome into the cytosol. The process described above produces a dipeptide bound to the tRNA. Now an elongation factor called translocase intervenes, which shifts the tRNA-bound dipeptide to the so-called donor (peptidyl) P site along with the entire mRNA chain, performing the translocation. The process repeats until a stop codon is placed on the small subunit. The information carried by the mRNA is read simultaneously by several ribosomes grouped along the mRNA strands called polysomes.

Antibiotics that inhibit protein synthesis affect the peptide chain elongation process by various mechanisms.

Inhibition of nucleic acid synthesis

Quinolones, actinomycin, mitomycin (DNA), rifampicin (mRNA), trimethoprim, sulfonamides (k. folic acid) act through this route.

Evaluation of antimicrobial effect

The evaluation is performed in vitro by determining the **minimum inhibitory concentration (MIC)** and the **minimum bactericidal concentration (MBC)**.

The MIC is the smallest measured amount (concentration) of an antibiotic that inhibits the growth and multiplication of bacteria in the test medium. The MBC corresponds to the lowest measured concentration of the antibiotic in vitro that will kill the exposed bacterial culture within 24 hours. Both concentrations are determined for bactericidal and bacteriostatic agents. For strong-acting bactericidal antibiotics, the differences between MIC and MBC are small. Proof of the reliability of the bactericidal effect is given by a subsequent culture test on agar media where no bacterial colonies are allowed to grow.

In order for the in vivo effect of an antibiotic to be sufficient, the minimum concentrations of the antibiotic in the target tissues must be consistent with the MIC and MBC values. In some cases (aminoglycosides, quinolones) the intensity of the antibacterial effect is directly proportional to the plasma and target tissue concentrations. High concentrations are not beneficial for the therapeutic effect, on the contrary, they burden the patient with the risk of adverse and toxic reactions. For most antibiotics, the duration of exposure is important for the therapeutic effect.

An important feature of some antibiotics (aminoglycosides and β -lactams) is the **post-antibiotic effect (PAE)**. This refers to the period of time during which bacterial multiplication ceases under conditions where the bacteria are no longer exposed to the antibiotic (no measurable concentration of the antibiotic in body fluids or tissues). The nature of this phenomenon has not yet been reliably explained.

Risks of antibiotic therapy

One of the serious risks of antibiotic administration is the development of adverse and **toxic effects** of antibiotics.

Adverse and toxic effects

Adverse effects occur at usual doses and recommended pharmacotherapeutic plasma concentrations. Toxic effects occur after high doses, due to high plasma concentrations, possibly with increased patient sensitivity. They can usually be avoided or their clinical manifestations mitigated. The risk of toxic effects is acceptable in the presence of life-threatening diseases, unless another, safer antibiotic is available.

Resistance

Another risk is the development of resistance (resistance) of microorganisms to the antibiotic.

Principles of antimicrobial use

The policy on the use of antimicrobials relates to:

1. **the correct choice of antimicrobial,**
2. **optimal and sufficient duration of treatment,**
3. **adequate dosage,**
4. **appropriate drug combinations,**
5. **desirable monitoring of antimicrobial therapy.**

The right choice of antimicrobial

The treatment of **chronic infections** is based on a **correct diagnosis and identification of the infectious agent**. This procedure allows for so-called targeted antibiotic therapy. First, the causative organism is isolated and then its susceptibility to antibiotics is determined. From the possible active substances, the one with the narrowest spectrum of action and the lowest possible toxicity (penicillin G in the case of pneumococcal infection) is selected. **Chronic infections tend to be caused by highly resistant agents (pseudomonads or staphylococci).**

In **acute infections**, targeted antibiotic therapy is rather the exception. In these cases, we usually choose **empirical treatment** (without bacteriological evidence of the causative agent). Empirical treatment consists of choosing the antibiotic that most closely matches the expected spectrum of pathogenic microorganisms and has the necessary pharmacokinetic properties. For example, different antibiotics are chosen for the treatment of wound infections, usually caused by *staphylococci* or *streptococci*, than for the treatment of urinary tract infections, usually caused by enterobacteriaceae, where the antibiotic needs to be renally excreted and reach high concentrations in the target tissue.

For **life-threatening infections**, where it is necessary to induce a sufficient effect as quickly as possible without knowing the causative agent, **broad-spectrum antibiotics (imipenem, ciprofloxacin) or combinations of antibiotics that together cover a broad spectrum (cefotaxime + piperacillin) are indicated**. This procedure is used for sepsis, peritonitis, infected gangrene, extensive infected burns, dangerous postoperative infections, endometritis, adnexitis and febrile abortion. Knowledge of the underlying or intercurrent disease is equally important for the correct choice of antimicrobial agent. It is necessary to search for risk factors, especially history of allergic reactions. Changes in the function of the eliminating organs due to pathology or abnormal conditions, e.g. low or high age, immune status, etc., are important. We relate this information to the pharmacological properties of the antimicrobial substance, i.e. its pharmacokinetics and pharmacodynamics (in particular potential adverse and toxic effects).

The ever-expanding range of antibiotics and the lack of insight into their therapeutic benefits tempt physicians to prescribe drugs with an unnecessarily broad spectrum and potent effect. In addition to the development of **multidrug resistance**, this trend has also led to a substantial increase in financial costs. A more fundamental change should be the introduction of a categorisation of antibiotics into groups: a - essential = essential b - alternative c - reserve and the development of binding treatment guidelines for specific diseases.

Treatment period

It is **based on the knowledge of the causative agent, the localization of the infection and the condition of the patient**. Usually, continuous treatment for **7-10 days** is sufficient. Therapeutic success is demonstrated by a decline in clinical and laboratory signs of infection (mainly temperature). However, in some **chronic processes** (osteomyelitis, tuberculosis, etc.), **antibiotic therapy may take months**. In an effort to reduce the risk of adverse effects, improve patient cooperation (compliance) and reduce the cost of therapy, single-dose regimens have been developed for the treatment of minor, uncomplicated infections (e.g. gonorrhoea, lues). For this purpose, bactericidal antibiotics with a longer $t_{1/2}$ (e.g. ceftriaxone, ciprofloxacin) or antibiotics with a sufficiently

long postantibiotic effect (spectinomycin in gonorrhoea) are suitable. A perfect proof of efficacy and safety of this procedure is a prerequisite. Prematurely discontinued antimicrobial therapy is one of the main factors involved in the emergence of resistance in bacterial strains.

Dose size

We distinguish between **single dose** (dosis singula, pro dosi), **daily dose** (dosis pro die) and **total therapeutic dose** (dosis curativa). The size of the dose is determined by the nature of the infection and the properties of the antibiotic (its pharmacokinetics and pharmacodynamics), the current condition of the patient (determined mainly by the function of the eliminating organs), the age of the patient and the route of administration. In clinical practice, the dosage recommended by the manufacturer is usually used, but may be influenced by the commercial interests of the company. For this reason, recommendations from independent clinical studies are considered to be the most relevant.

Dosage adjustments can be made according to two principles:

- **pharmacodynamic principle** - for antibiotics with a wide therapeutic range (interval between the minimum toxic dose and the minimum therapeutic dose), the dose size can be modified according to the severity of the disease.
- **pharmacokinetic principle** - for antibiotics with a narrow therapeutic range (aminoglycosides), increasing the dose is risky. The dose size should be adjusted according to plasma concentrations.

Route of administration

The route of administration depends on the pharmacokinetic properties of the antibiotic, i.e. its absorption, distribution and elimination, as well as the patient's condition and the nature of the infection.

- **Parenteral** - administration in severe systemic infections (sepsis, meningitis) and where oral administration cannot achieve sufficient concentration at the site of infection or the patient's condition does not allow other than parenteral administration (unconsciousness, GIT disease).
- **Orally** - this antibiotic treatment may be local or total. Total therapy is for systemic effects and is intended for the treatment of mild and moderate infections. The basic prerequisite for its choice is sufficient bioavailability. In sequential therapy, the patient is switched to oral therapy (injectable cefuroxime to the esterified oral dosage form cefuroxime-axetil) after 2-3 days of injection therapy.

Combination of drugs

Combination of drugs implies the use of two or more antibiotics at the same time. It is indicated for several reasons:

- in the empirical treatment of serious infections (sepsis) to cover the full spectrum of possible causative agents. In these cases, a specific type of empirical therapy is chosen, namely, stepwise - interventional - treatment according to a specific regimen. The scheme specifies which additional antibiotics should be continued in the event of ineffective initial therapy. This procedure is usually used in the treatment of infectious diseases associated with impaired immune function, e.g. febrile neutropenia, secondary infection in AIDS patients, etc.
- The combination of antibiotics can achieve a significant enhancement of the effect (synergism) of both agents. The essence of synergism is:

1. sequential blockade - by simultaneous inhibition of successive metabolic processes essential for the life of the bacterium (trimethoprim + sulfonamide),

2. inhibition of enzymatic inactivation (amoxicillin with clavulanic acid),

3. increasing antibiotic penetration into the bacterium (combination of β -lactam antibiotics with aminoglycoside).

- The combination of antibiotics can prevent the rapid development of resistance where the administration of a single antibiotic is most likely to lead to the development of resistance (in tuberculosis).
- Finally, combinations may allow the doses of toxic antibiotics to be reduced while maintaining their effect.

Monitoring antimicrobial therapy

The basic principle of monitoring antimicrobial therapy is to monitor the clinical status of the patient. Above all, it is necessary to look for signs of therapy failure, for the development of adverse or toxic effects, which may be reversible in the first phase. Monitoring of plasma concentrations is recommended:

- antibiotics where there is a significant relationship between plasma concentrations and therapeutic or toxic effects,
- antibiotics with a narrow therapeutic window, i.e. with a small range between effective and toxic concentrations (aminoglycosides),
- in at-risk patient groups (chloramphenicol in newborns).

To ensure rational antibiotic pharmacotherapy, it is necessary to know the results of culture and resistance determination.

Failures and complications of antimicrobial therapy

It manifests itself clinically by persistence of temperature, pain, inflammatory manifestations or completely new symptomatology. The most common causes of failure are:

- the infectious agent is not sufficiently sensitive to the antibiotic, i.e. the drug was not selected correctly, the in vitro culture results were false positives, or there are differences between the in vitro and in vivo effect of the antibiotic,
- Inadequate levels at the site of infection with inappropriate choice of dosage form (p.o./i.v.), inadequate dosage or inadequate pharmacokinetic properties of the drug (substance does not penetrate barriers),
- drug interactions may cause a reduction in effect (reduced absorption of tetracyclines after administration of antacids, accelerated metabolism after rifampicin) or, conversely, induce toxic effects (furosemide increases the nephrotoxicity of gentamicin),
- toxic manifestations due to changes in the function of elimination organs, e.g. renal insufficiency, "gray syndrome" in newborns, etc.
- allergic reactions

The most commonly used abbreviations of antibiotics

Antibiotic	Abbreviation
Amikacin	AMI
Amoxicillin	AMO
Amoxicillin + clavulanic acid	AMC
Ampicillin	AMP
Ampicillin + sulbactam	AMS
Azithromycin	AZI
Azlocillin	AZL
Aztreonam	AZT
Bacitracin	BAC
Cefaclor	CCL
Cefadroxil	CDR
Cefalexin	CLX
Cephalothin	CLT
Cefamandole	CMN
Cefapirin	CPN
Cefazolin	CZL
Cefepim	CPM
Cefetamet pivoxil	CTP
Cefoperazone	CPR
Cefoperazone + sulbactam	CPS
Cefotaxime	CTX
Cefoxitin	CXT
Cefpirom	CRM
Cefpodoxime proxetil	CPD
Cefsulodin	CSL
Ceftazidime	CTZ
Ceftibuten	CTB
Ceftizoxime	CTM
Ceftriaxone	CTR
Cefuroxime	CRX
Ciprofloxacin	CIP
Doxycylin	DOX
Erytromycin	ERY

Ehambutol	ETM
Etionamide	ETI
Fusidic acid	FUS
Gentamicin	GEN
Chloramphenicol	CMP
Imipenem/cilastatin	IMI
Isoniazid	INH
Josamycin	JOS
Kanamycin	KAN
Clarithromycin	CLA
Clindamycin	CLI
Colistin	COL
Cotrimoxazole	COT
Linkomycin	LIN
Lomefloxacin	LOM
Meropenem	MEM
Metronidazole	MET
Neomycin	NEO
Netilmicin	NET
Nitrofurantoin	FUR
Norfloxacin	NOR
Ofloxacin	OFL
Oxacilin	OXA
Oxalic acid	OXO
Pefloxacin	PEF
Penicillin	PEN
Piperacillin	PIP
Piperacillin/tazobactam	PPT
Polymyxin	POL
Pyrazinamide	PYR
Rifabutin	RBT
Rifampicin	RIF
Roxithromycin	ROX
Sparfloxacin	SPR
Spektinomycin	AKS
Spiramycin	SPI
Streptomycin	STR
Sulfonamides	SUL
Teikoplanin	TEI
Tetracycline	TET
Tikarcillin	TIC
Tikarcillin+klavulanic acid	TIM
Tobramycin	TOB
Trimethoprim	TRI
Trovafloxacin	TRV
Vankomycin	VAN

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Antibiotics and chemoterapeutics

baktericidal	aminoglycosides	amikacin, gentamycin, isepamycin, neomycin, netilmicin, spectinomycin, streptomycin, tobramycin			
	antituberkulotics	izoniazid, cykloserin, ethambutol, ethionamid, kapreomycin, pyrazinamid, viomycin			
	beta-lactam	cephalosporins	I.generation	cafazolin, cefadroxil, cefalexin, cefalotin, cefapirin	
			II.generation	cefuroxim, cefamandol, cefpodoxim, proxetil, cefprozil monohydrate, cefuroxim-axetil	
			III.generation	cefotaxim, cefetamet pivoxil, cefixim, cefoperazon, cefsulodin, ceftazidim, ceftibuten, ceftriaxon, co-cefoperazon	
			IV.generation	cefepim, ceftiprom	
			V.generation	ceftarolin	
		carbapenems	imipenem, doripenem, ertapenem, merapenem		
		monobaktams	aztreonam		
		penicillins	narrow-spectrum penicillins	penicillin G, penicillin V, cloxacilin, dicloxacilin, oxacilin, flucloxacilin	
			broad-spectrum penicillins	amoxicilin, ampicilin, ticarcilin, piperacilin	
			beta-lactam inhibitors	co-amoxicilin, co-ticarcilin, sulbaktam, tazobaktam	
	glycopeptides	teikoplanin, vankomycin			
	quinolones	I.generation	nalidixic acid, oxolinic acid		
		II.generation	ciprofloxacin, norfloxacin, pipemidic acid, rosoxacin		
		III.generation	sparfloxacin, enoxacin, fleroxacin, lomefloxacin, ofloxacin, pefloxacin		
		IV.generation	moxifloxacin		
	nitroimidazoles	metronidazol, ornidazol			
	nitrofurans	furazolidon, nifuratel, nifuroxazid, nitrofurantoin,			
polypeptide antibiotics	bacitracin, colistin, gramicidin, polymyxin B				
rifamycins	rifampicin, rifabutin, rifaximin, rifamycin				
bakteriostatic	amphenicol	azidamfenikol, chloramfenikol, florfenikol, thiamfenikol			
	glycylcyclines	tigecyclin			
	lincosamides	klindamycin, linkomycin			
	macrolides	I.generation	erytromycin, josamycin, oleandomycin, spiramycin		
		II.generation	azithromycin, clarithromycin, roxithromycin		
	oxazolidinones	linezolid			
	pyrimidines	co-trimoxazol, pyrimetamin, trimetoprim			
	streptogramins	streptogramin A, B			
	sulphonamides	sulfamothoxazol, sulfasalazin, sulfathiazol, sulfisoxazol			
	sulfones	dapson			
tetracyclines	doxycyklin				